Alternaria species: endophytic fungi as alternative sources of bioactive compounds

Daniya Eram¹, Manojkumar Arthikala², Govindappa Melappa¹ and Gustavo

Santoyo³

¹Department of Biotechnology, Dayananda Sagar College of Engineering, Shavige Malleshwara Hills, Kumaraswamy Layout, Bengaluru-560 078, Karnataka, India. ²Ciencias Agrogenómicas, Escuela Nacional de Estudios Superiores, León, Universidad Nacional Autónoma de México (UNAM), C.P.-37684, Mexico.

³Instituto de Investigaciones Químico-Biológicas, Universidad Michoacana de San Nicolás de Hidalgo, Morelia, Michoacán, Mexico

Correspondig Author e-mail: dravidateja07@gmail.com, drgovindappa-bt@dayanandasagar.edu

Abstract

Fungal endophytes are a rich source of functional metabolites. In particular, the different species of the genus *Alternaria* stand out for their arsenal of metabolites, which have roles as antimicrobial molecules, antioxidants; as well as activities against HIV or cancer or diabetes. This review will highlight on selected aspects of the fungal endophytes of *Alternaria* species: their widespread distribution, and the role of their metabolites with pharmaceutical or agronomical importance. The family plants associated with *Alternaria* species, their bioactive compounds and biological activity to date are examined.

Keywords: Alternaria species; metabolites; antimicrobials; cancer; diabetes; HIV

Riassunto

I funghi endofiti delle piante sono ricche fonti di metaboliti bioattivi. In particolare le specie appartenenti al gen. *Alternaria* producono una serie di molecole ad azione antimicrobica, antiossidante, anti HIV, antitumorale e antidiabetica. Nel presente lavoro vengono esaminate le piante associate alle specie di *Alternaria* e vengono riportati i composti bioattivi prodotti e la loro attività.

Parole chiave: Alternaria spp.; metaboliti; azione antimicrobica; cancro; diabete; HIV

Introduction

Endophytes are the micro-organisms that colonize the inside of the plant parts, without having any negative impact on the host (Arnold *et al.*, 2003), and these contribute resistance against both biotic (Breen, 1994; Schulz *et al.*, 1999; Dingle and McGee, 2003) and abiotic stresses (Siegel *et al.*, 1990; West, 1994). The researchers found endophytic fungi in almost all studied plants and thus it implies an important component of plant micro-ecosystem (Strobel, 2006). In addition, the

endophytes impart resistance against plant pathogens and herbivores and they are also able to produce known or unknown pharmaceutical and agricultural important bioactive compounds. Most of endophytic organism bioactive compounds act as plant defence molecules and some of them are using to treat various human diseases (Owen and Hundley, 2004). Endophytic fungi from Muscador albus of rainforest plant are able to produce fumigant agent against stored grain pests (Strobel, 2006; Strobel et al., 2001). The endophytic fungi are able to produce many antimicrobial metabolites, such as colletotric acid (Zou et al., 2000) and griseofulvin (Park et al., 2005) reported from Alternaria species ubiquitous fungal genus including saprobic, endophytic and pathogenic species. Alternaria species are associated with a wide variety of substrates including seeds, plants, agricultural products, animals, soil and the atmosphere. Pathogenic species of Alternaria pose variety of phytopathological problems which results in yield losses in a wide range of crop plants. Several taxa are also important postharvest pathogens, causative agents of phaeohyphomycosis in immuno-compromised patients or airborne allergens. von Esenbeck (1816) originally described Alternaria, based on A. tenuis as the only species. Characteristics of the genus included the production of dark-coloured phaeodictyospores in chains, and a beak of tapering apical cells. von Keissler (1912) synonymised both A. tenuis and Torula alternata (Fries, 1832) with Alternaria alternata, due to ambiguities in Nees's description of A. tenuis. Two additional genera, Stemphylium (Wallroth, 1833) and Ulocladium (Preuss, 1851) were subsequently described for phaeodictyosporichyphomycetes, further complicating the taxonomic resolution in this group of fungi. Several re-descriptions and revised criteria of these genera (Saccardo, 1886; Elliot 1917; Wiltshire, 1933, 1938; Joly, 1964) resulted in a growing number of new species. Results of a lifetime study on Alternaria taxonomy based on morphological characteristics were summarised by Simmons (2007), in which 275 Alternaria species were recognised. One species was transferred to the genus Prathoda and three new genera, Alternariaster, Chalastospora and Teretispora were segregated from genus Alternaria. Molecular studies revealed multiple non-monophyletic genera within the Alternaria complex and Alternaria species clades, however based on morpholigical characteristics it do not correlate to species-groups (de Hoog and Horré, 2002; Pryor and Bigelow, 2003; Hong et al., 2005; Inderbitzin et al., 2006; Pryor et al. 2009; Runa et al., 2009; Lawrence et al., 2012). The A. alternata, A. brassicicola, A. infectoria, A. porri and A. radicina species groups were strongly supported by these studies and two new species-groups, A. sonchi (Hong et al., 2005) and A. alternantherae (Lawrence et al., 2012) and two new genera, Crivellia (Inderbitzin et al., 2006), Undifilum (Pryor et al., 2009) were described. The latest molecular revision of Alternaria (Lawrence et al., 2012) introduced two new species groups, A. panax and A. gypsophilae, and elevated eight species-groups to sections within Alternaria. The sexual phylogenetic Alternaria lineage, the A. infectoria species-group, did not get the status of section, in contrast to the eight asexual phylogenetic lineages in Alternaria. The Alternaria complex currently comprises the genera Alternaria, Chalastospora (Simmons, 2007), Crivellia, Embellisia, Nimbya, Stemphylium, Ulocladium, Undifilum and the recently described Sinomyces together with eight sections of Alternaria and the A. infectoria species-group. The present review focused on endophytic fungi, Alternaria species, their phytochemicals, biological activity and their plant hosts in detail.

Spreading family

Details of endophytic *Alternaria* species, hosts, natural compounds are represented in table 1. Their biological activities are briefly explained in figure 1. Endophytic fungi, *Alternaria* species been found in various plant species belonging to different families. *Alternaria* species in *Taxus cuspidata, T. yunnanensis* and *T. chinensis var. mairei* belonging to the *Taxaceae* family produce taxol, cladosporol

and paclitaxel which mainly exhibit anti-cancer/ anti-tumour properties (Vasundhara *et al.*, 2016; Chen *et al.*, 2009; Strobel *et al.*, 1996; Kim and Ford, 1999; Tian *et al.*, 2006; Zhou *et al.*, 2007; Guo *et al.*, 2009; Wu *et al.*, 2011). Li and co-workers (2016a) obtained 940 isolates from different tissues of *Zanthoxylum bungeanum* and were grouped into 93 morphotypes, 43 species, and 23 genera. Potent antioxidant and antimicrobial activity was observed with extract of *A. alternata*, isolated from bark of *Aegle marmelos* (Mani *et al.*, 2015; Selvi and Balagengatharathilagam, 2014) and *Phellodendron amurense* (Duan, 2009) belonging to *Rutaceae* family.

Alternaria alternata (isolated from *Eugenia jambolana*) extract exhibited strong antibacterial property against MDR (Multi-Drug Resistance) (Yadav *et al.*, 2016). *Salvia miltiorrhiza* (Tian *et al.*, 2017; Lou *et al.*, 2016), *Salvia przewalskii* (Wang *et al.*, 2014) and *Ocimum sanctum* (Chowdhary and Kaushik, 2015) of *Lamiaceae* family have exhibited *Alternaria* species and it showed anti-bacterial and anti-phytopathogenic property.

Alternaria tenuissima was reported from bark of *Erythrophleum fordii* (Fang *et al.*, 2012) able to produce cyclopeptides, and also from leaves of *Acacia magnum* produce alternariol monomethyl ether and α , β - dehydrocurvularin. These biomolecules has been reported to have inhibitory effect on *Magnaporthe grisea* appressorium development (Jeon *et al.*, 2010).

Alternaria brassicola isolated from *Malus halliana* (*Rosaceae*) producing alternariol, alternariol 9methyl ether, altechromone A, herbarin A, cerevisterol, 3β , 5α - 22E, 24R)-3,5-dihydroxyergosta-7,22dien-6-one and 3β , 5α -dihydroxy- (22E,24R)-ergosta-5,8,22-trien-7-one are antimicrobial agents (Guo *et al.*, 2009). A phenylpropene known as eugenol was identified in *Alternaria* species isolated from *Rosa damacaena* (Rosaceae) (Kaul *et al.*, 2008). Anti-insecticidal podophyllotoxin was noticed in extract of *Alternaria* species isolated from *Sabina vulgaris* (*Cupressaceae*) (Lu *et al.*, 2006).

The *Alternaria* species was isolated from root of marine semi-mangrove plant, *Myoporum bontioides* (*Scrophulariaceae*) able to produce cyclohexenone, cyclopentenone and xanthone and were reported to be a strong antioxidant and antimicrobial agents (Wang *et al.*, 2014). Antibacterial agent altenusin was noticed in the extract of *Alternaria* species, endophytic fungi of *Sonneratia alba* (*Lythraceae*) (Kjer *et al.*, 2009).

Endophyte, *Alternaria* species was also isolated from *Ginkgo biloba* of *Ginkoaceae* (Qin *et al.*, 2009) and extracts showed strong antifungal activity (Xiao *et al.*, 2013). *Alternaria* species isolated from phloem of *Catharanthus roseus* (*Apocynaceae*) had the ability to produce vinblastine (Zhang *et al.*, 1998). Podophyllotoxin was identified from *Alternaria neesex* of *Sinopodophyllum hexandrum* (*Berberidaceae*) (Li, 2007; Cao *et al.*, 2007; Yang *et al.*, 2003) and also from *Alternaria* species obtained from *Sabina vulgaris* (Lu *et al.*, 2006). Paclitaxel was isolated from *Alternaria* species (*Taxus cuspidata*), *Alternaria* species (*Ginkgo biloba*), *Alternaria alternata* (*Taxus chinensis* var *mairei*) (Strobel *et al.*, 1996; Kim *et al.*, 1999; Tian *et al.*, 2006).

The endophytic Alternaria species were also reported from *Quercus emoryi* of *Fagaceae* (Bashyal *et al.*, 2014), *Tabebuia argentea* of *Bignoniaceae* (Channabasava and Govindappa, 2014), *Viscum album* of *Santalaceae* (Govindappa *et al.*, 2015), *Trixis vauthieri* of *Asteraceae* (Cota *et al.*, 2008), *Ficus carica* of *Moraceae* (Feng and Ma, 2010), *Denderonephthya hemprichi* of *Nephtheidaceae* (Shaaban *et al.*, 2012), *Polygonum senegalense* of *Polygonaceae* (Aly *et al.*, 2008), *Azadirachta indica* of *Meliaceae* (Taware and Rajurkar, 2015), *Aegiceras corniculatum* of *Primulaceae* (Huang *et al.*, 2011), *Laurencia* species of *Rhodomelaceae* (Gao *et al.*, 2009), *Paeonia delavayi* of *Paeoniaceae* (Wu *et al.*, 2011), *Maytenus hookeri* of *Celastraceae* (Yuan *et al.*, 2008), *Tylophora indica* of *Apocynaceae* (Kumar *et al.*, 2011) and *Vitis vinifera* of *Vitaceae* family (Shi *et al.*, 2012).

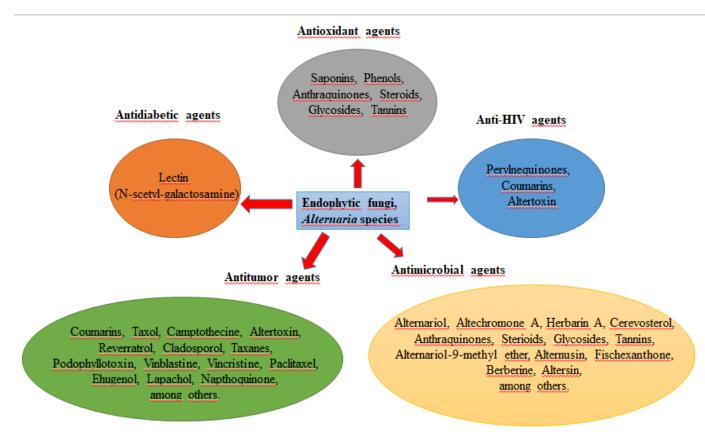


Fig. 1. Important phytochemicals from endophytic fungi, *Alternaria* species and their biological activity Fig. 1. Importanti prodotti fitochimici da specie di *Alternaria* e loro attività biologica.

Antimicrobial molecules

Alternaria species or A. alternata from Camptotheca acuminata (Ding et al., 2006), Achyranthus aspera, Adhatoda zeylanica, Aegle marmelos, Leucas aspera, Azadirachta indica (Selvi and Balagengatharathilagam, 2014), A. raphami from Vitax negundo (Monali et al., 2013), A.tenussima from Ricinus communis (Sandhu et al., 2014a) were the promising antibacterial compounds against different pathogenic bacterial species.

Extracts of *Alternaria* species isolates of *Tephrosia purpurea* (Luo *et al.*, 2015) and *Opuntia humifusa* (Silva-Hughes, 2015), *A. alternata* from *Suaeda martiana, Sueada monoica* (Kalyanasundaram *et al.*, 2015), *A. solani* from *Heptacdium miconioides* (Wang *et al.*, 2016), *Altrnaria* species had shown strong antifungal activity against plant pathogens. The *Alternaria species* of *Garcinia* plant exhibited antiviral activity (Wiyakrutta *et al.*, 2004; Phongpaichit *et al.*, 2007). *Alternaria* species from *Mllotus philippinensis* (Gangwar *et al.*, 2015), *Loranthus* (Govindappa *et al.*, 2011), *Artemissia* species (Cosoveanu *et al.*, 2016) extract showed strong antimicrobial activity.

Antioxidant molecules

Oxidative stress has been identified as the root cause of the development and progression of several diseases. Supplementation of exogenous antioxidants or boosting endogenous antioxidant defences of the body is a promising way of combating the undesirable effects of reactive oxygen species (ROS) induced oxidative damage. Many medicinal plants contain large amounts of antioxidants other than vitamin C, E and carotenoids (Javanmardi *et al.*, 2003). Antioxidants are molecules that can delay or

prevent an oxidative reaction (Velioglu *et al.*, 1998) catalysed by free radicals. This antioxidant effect is mainly due to the presence of phenolic components such as flavonoids (Pietta *et al.*, 1998), phenolic acids and phenolic diterpenes (Shahidi *et al.*, 1992). Antioxidants such as Butylated Hydroxy-Anisol (BHA), Butylated Hydroxy-Toluene (BHT) protect plants against oxidative assault (Dziezak, 1986) either by binding to metallic ions, eliminating free radicals or by decomposing peroxides (Matook, 2005). Despite the availability of synthetic antioxidants, present research seeks at discovering new natural antioxidant compounds that may play a role in oxidative stress related disorders (Agbor *et al.*, 2005). Epidemiological studies (Urquiaga and Leighton, 2000; Halliwell, 1994) have showed that decreases in the incidence of chronic diseases in some populations were related to the consumption of fruits and vegetables. Antioxidants such as saponins, phenolic compounds, anthraquinones, steroids, cardiac glycosides and tannins are produced by endophytic *Alternaria* species found in *Aegle marmelos* (Selvi and Balagengatharathilagam, 2014) and Dibenzo- α -pyrones (1–7) in *Salvia miltiorrhiza* Bunge (Tian *et al.*, 2017). The crude extracts of *A. alternata* from *Coffea arabica* (Fernandes *et al.*, 2009), *Alternaria* species from *Mussaenda luteola* (Gunasekharan *et al.*, 2017) showed strong antioxidant activity.

Anti-diabetic molecules

In humans, diabetes mellitus is considered as one of the five leading causes of death in the world (Joseph and Jini, 2011). Diabetes mellitus is a major global health concerning with a projected rise in prevalence from 171 million in 2000 to 366 million in 2030 (Shaw et al., 2010). It is a syndrome of disordered metabolism, usually due to a combination of hereditary and environmental causes, resulting in abnormally high blood sugar levels (hyperglycemia) (Patel et al., 2012). Being a major degenerative disease, diabetes is found in all parts of the world and increasing rapidly and thus it is the third most lethal disease of mankind (Ogbonnia et al., 2008). It is the most common endocrine disorder, affecting 16 million individuals in the United States alone and as many as 200 million individuals worldwide. Diabetes has been a clinical model for general medicine (Sharma et al., 2012). Complementary and alternative medicine involves the use of herbs and other dietary supplements as alternatives to mainstream western medical treatment. A recent study has estimated that up to 30% of patients with diabetes mellitus use complementary and alternative medicine (Raman et al., 2012). Alternaria species can be an excellent source for production of anti-diabetic molecules. Lectin (Nacetyl-Galactosamine) produced by endophytic Alternaria species of Viscum album been characterized to process has anti-diabetic properties (Govindappa et al., 2015). In another study, Alternaria species from Phyllanthus emblica fruits (Singh et al., 2017) and Morus alba (Zheng et al., 2014) exhibited strong inhibitory effect on diabetic enzymes.

Anti-HIV molecules

Since the first cases of AIDS were identified in 1981 in the United States. AIDS has become the largest and most devastating public health pandemic of our time, and has infected nearly 70 million people and left 25 million dead, and around the world, the number of people living with HIV is about 36.7 million. Combination therapy of anti-HIV drugs now available has improved the quality of life and lifespan of HIV/AIDS patients. Emergence of HIV drug resistance, side effects and the need for long-term antiretro-viral treatment are the main causes for the failure of anti-retroviral therapy (ART). Continuous development of new anti-HIV agents, targets and therapy appear to be inevitable. In some countries, traditional medicine is used to meet primary health care needs and to treat AIDS patients (Harnett *et al.*, 2005). Natural products are the most consistently successful source in drug discovery, and may offer more opportunities to find anti-HIV drugs or lead compounds. Many compounds with an anti-HIV-1 effect have been screened out from natural products and discovered to inhibit HIV at nearly all stages of the viral life cycle (Wang *et al.*, 2004). They include alkaloids, sulphated polysaccharides, polyphenolics, flavonoids, coumarins, phenolic, tannins, triterpenes, lectins, phloroglucinols, lactones, iridoids, depsidones, O-caffeoyl derivatives, lignans, ribosome-inactivating proteins, saponins, xanthones, naph-thadianthrones, photosensitisers, phosholipids, quinines and peptides (Ng *et al.*, 1997; Vlietinck *et al.*, 1998; Yang *et al.*, 2001). Natural products provide a large reservoir for screening anti-HIV agents with novel structure and anti-viral mechanisms because of their structural diversity. A variety of natural products have been found to inhibit unique enzymes and proteins crucial to the life cycle of HIV, including efficient intervention with the reverse transcription process, virus entry, integrase and protease (De Clercq, 2000; Cos *et al.*, 2004). But the mechanism of anti-HIV activity of many more natural products is still unknown.

People living with HIV/AIDS often choose traditional or complementary and alternative medicine to complement or replace conventional treatment. The presence of multi-drug or even multiclass resistance in HIV also warrants the need to explore additional means to combat HIV and provide further justifications for the need of alternative and complementary medicines in the treatment of HIV/ AIDS. Most of the traditional systems of medicine in India include some form of 'medicinal plant', herbs or natural plant products. It is therefore not surprising that the activity of these traditional medicines against HIV can be scientifically analysed to deduce the role of natural plant products in their anti-HIV activities. A number of medicinal plants have been reported to have anti-HIV properties (Fritts *et al.*, 2008). Over the past two decades, substantial progress has been made in research on the natural products possessing anti-HIV activity. A variety of secondary metabolites obtained from natural origin showed moderate to good anti-HIV activity.

Endophytes present in various plants can also be a potential source for production of anti-HIV molecules (Singh *et al.*, 2005). Altertoxins produced by *A. tenussima* shows a significant anti-HIV activity (Bashyal *et al.*, 2014; Wellensiek *et al.*, 2013). Partially purified coumarin extracts of *Alternaria* species from *Calophyllum inophyllum*) have inhibited the HIV-1 replicating proteins significantly (Govindappa *et al.*, 2015). Perylnequinones isolated from *A. tenussima* of *Quercus emoryi* have shown potent HIV-1 inhibitors (Hudson *et al.*, 1997).

Anti-microbial molecules

Endophytes, microorganism that reside in the tissue of living plants, are relatively understudied and potential source of novel natural products for exploitation in medicine, agriculture and industries (Sandhu *et al.*, 2014b). The resistance of pathogenic microorganisms to drugs and antibiotics has become a major challenge in the health sector leading to reduction in drug effectiveness and economic wastage. This challenge has once again stirred up a need in scientific research and the discovery of new more effective antimicrobial metabolites becoming a major research interest. In recent years, the isolation of endophytic fungi and screening of antimicrobial activity has gained more attention (Strobel, 2003).

The increase in the resistance of drugs by infectious pathogens as well as undesirable effects of certain antimicrobial agents indicates that there is an urgent need for novel and effective bioactive compounds with fresh modes of action which is what this study is aimed at achieving stated that during the last 20 years (Strobel, 2006; Porras-Alfaro and Bayman, 2011). It has been observed that much of the wealth of microbial biodiversity with novel biochemistry and secondary metabolite production resides in plant tissues. The endophytes of these plants, especially the *Alternaria* species produce a wide range of anti-microbial compounds.

Fischexanthone, an anti-fungal compound, is produced by endophytic *Alternaria* from root tissues of *Myoporum bontioides* (Li *et al.*, 2016b). Strong anti-fungal activity was observed in *Ficus carica* due to the production of helvolic acid by the endophytic *Alternaria* (Feng and Ma, 2010). Cyclo-[L-Leu-trans-4-hydroxy-L-Pro-], Cyclo-(L- Phe-trans-4-hydroxy-L-Pro-), and Musetti *et al.* (2007) have produced Cyclo-(L-Ala-trans-4-hydroxy-L-Pro) in broth culture of the grapevine also exhibits anti-fungal activity. Xanalteric acids I and II produced in *Somneratia alba* exhibits weak anti-bacterial activity against multidrug-resistant *Staphylococcus aureus* (Kjer *et al.*, 2009). Solanapyrones P–R (1–3), solanapyrones (4–6) and benzopyrones (7–9) produced in the root tissues of a Chinese herbal medicinal plant *Salvia przewalskii* also exhibits anti-bacterial activity (Xiao *et al.*, 2014). Apart from these dibenzo- α -pyrones produced in *Salvia miltiorrhiza* (Danshan) due to endophytic *Alternaria* also shows anti-bacterial activity (Tian *et al.*, 2017).

Altersin produced by Alternaria shows broad anti-microbial activity (Hellwig et al., 2002; Fernandes et al., 2009). Alternusin produced by endophytic Alternaria species of Sonneratia alba exhibits antimicrobial activity against several additional multidrug-resistant bacterial and fungal strains (Kjer et al., 2009). Alternariol 9-methyl ether (AME) in Salvia miltiorrhiza (Lou et al., 2016); berberine in Phellodendron amurense (Duan, 2009); saponins, phenolic compounds, anthraquinones, steroids, cardiac glycosides and tannins in Aegle marmelos (Selvi and Balagengatharathilagam, 2014); pyrophen, rubrofusarin B, fonsecin, fonsecin B aurasperone A, aurasperone B, aurasperone C, and aurasperone F in Denderonephthya hemprichi (Shaaban et al., 2012); 7-epi-8-hydroxyaltertoxin I and 6-epi-stemphytriol, stemphyperylenol and altertoxin I in Laurencia sp. (Gao et al., 2009); alternariol, alternariol 9-methyl ether, altechromone A, herbarin A, cerevisterol, 3β,5α-dihydroxy-(22E,24R)ergosta-7,22-dien-6-one and 3β-hydroxy-(22E,24R)-ergosta-5,8,22-trien-7-one in Mallus haliana (Guo et al., 2009); alternariol, djalonensone, cerebroside B, cerebroside C, (2S, 3S, 4R, 2' R) -(2'hydroxytetracosanoylamino) octadecane-1,3,4-triol, and mannitol in Paeonia delavayi (Wu et al., 2011) were reported to process anti-microbial activity. Gunasekharan et al. (2017), Srinivasan et al. (2015) have reported that Alternaria species extracts have inhibited the activity of pathogenic bacteria. Two new antimicrobial compounds, 10-oxo-10H-phenaleno[1,2,3-de] chromene-2-carboxylic acids, xanalteric acids I and II and 11 known secondary metabolites were obtained from Alternaria species extract (Sonneratia alba) and they are strong antibacterial compounds (Kjer et al., 2009). The potent antifungal activity was observed from Alternaria species extract isolated from Clerodendrum *inerme* (Gong *et al.*, 2015).

Anti-cancer molecules

Cancer is considered one of the most common causes of human mortality worldwide. Progress made in cancer therapy has not been sufficient to a significantly lower annual death rate from most tumor types, and there is an urgent need for new strategies in cancer control (Mesnah *et al.*, 1987). For centuries, people have been using plants for their therapeutic values. Today 85000 plants have been documented for therapeutic use globally (Liu and Wang, 2008). The World Health Organization (WHO) estimates that almost 75% of World's population has therapeutic experience with herbal drugs. Cancer is one of the most widespread diseases in humans and presently there is a considerable scientific discovery of new anticancer agents from natural products (Kasabana and Hemini, 1998). The potential of using the natural products as anticancer drugs was recognized in 1950's by U.S. Natural Cancer Institute (NCI) since 1950 major contributions have taken for the discovery of naturally occurring anticancer drugs (Cragg and Newman, 2005).

In this sense, altertoxin IV showed significant anticancer activity was isolated from endophytic fungi *Alternaria* species of *Broussonetia papyrifera* (Zhang *et al.*, 2016). Taxol is producing by *Alternaria*

species of *Salacia oblonga* (Roopa *et al.*, 2015), *Taxus cuspidata* (Strobel *et al.*, 1996), *Ginkgo biloba* (Kim *et al.*, 1999), *Corylus avellena* (Michalczyk *et al.*, 2014; Soliman and Raizada, 2013) and paclitataxel from *Alternaria tenuissima* (Ismaiel *et al.*, 2017). Taxanes was isolated from *Alternaria* species of *Taxus baccata* (Staniek *et al.*, 2010). *Alternaria* species have ability to produce paclitaxel or its analogues (Zhao *et al.*, 2012; Chandra, 2012). Camptothecine (CPT) and its analogues 9-methoxycmptothecine (MCPT) and 10-hydroxycamptothecine (HCPT) produced by *Alternaria alternata* from *Miquelia dentata* (Shweta *et al.*, 2013). Anticancer agent, coumarins were reported from *Alternaria* species and *A. alternata* (Sadananda *et al.*, 2011; Channabasava and Govindappa, 2014), podophyllotoxin from *Alternaria* species (*S. hexandrum*). Vinblastine and vincristine were produced from *Alternaria* species isolated from *Catharanthus roseus* (Guo and Kunming, 1998). Kaul *et al.* (2008) have reported the eugenol produced by *Alternaria* species of rose. Reveratrol isolated from *Alternaria* species of *Vitis vinifera*, *Vitis quinquangularis*, *Polygonum cuspidatum* (Shi *et al.*, 2012) and cytotoxic compounds were isolated from *Alternaria* species of *Polygonum senegalense* (Aly *et al.*, 2008). *Salvia*

Discussion and future research

Alternaria species are an abundant source of bioactive molecules, with promising roles to fight against pathogenic microbes or human diseases, such as diabetes, HIV and cancer. However, it is suggested that the work done in the future include a step to take a step forward to apply all these promising metabolites. It is known that the regulations in each country of the world are different for the clinical application of new compounds, but undoubtedly, researchers from each region should make an effort to achieve this goal for the benefit of humanity.

miltiorrhiza (Lou *et al.*, 2016) exhibit anti-cancer activity. *Alternaria alternata* (*Cissus quadrangularis, Asclepias curassavica*) extract had shown strong cytotoxicity activity (Tenguria and Firodiya, 2016).

Likewise, attention is needed within the basic research on different compounds that produce the endophytic fungi, *Alternaria* species and to identify environmental aspects that regulate their production, as well as their effectiveness when applied. The aspects of resistance in antimicrobial compounds are also an area, requires a continuous research, because new resistance mechanisms arise with new drugs. Large scale commercial productions of such biomolecules using biotechnological tools are also in urgent used. Therefore, it is a potential area for investigation and pharmaceutical applications.

References

- Agbor A.G., Oben J.E., Ngogang J.Y., Xinxing C., Vinson J.A. (2005). Antioxidant Capacity of Some herbs/spices from Cameroon. a comparative study of two methods. Journal of Agriculture, Food Chemistry, 53(17):6819–6824.
- Aly A.H., Edrada-Ebel R.A., Indriani I.D., Wray V., Muller W.E.G., Totzke F., Zirrgiebel U., Schachtele C., Kubbutat M.H.G., Lin W.H., Proksch P., Ebel R. (2008). Cytotoxic metabolites from the fungal endophyte *Alternaria* sp. and their subsequent detection in its host plant *Polygonum senegalense*. Journal of Natural Products, 71: 972-980.
- Arnold A.E., Mejia L.C., Kyllo D., Rojas E.I., Maynard Z., Robbins N., Herre E.A. (2003). Fungal endophyte limit pathogen damage in a tropical tree. Proceedings of National Academy of Sciences USA, 100:15649–15654.
- Bashyal B.P., Wellensiek B.P., Ramakrishnan R., Faeth S.H., Ahmad N., Gunatilaka A.L.L. (2014). Altertoxins with potent anti-HIV activity from *Alternaria tenuissima* QUE1Se, a fungal endophyte of *Quercus emoryi*. Bioorganic & Medicinal Chemistry, 22:6112–6116.

- Breen J.P.(1994). Acremonium endophyte interactions with enhanced plant resistance to insects. Annual Review on Entomology, 39:401–423.
- Cao L., Huang J., Li J. (2007). Fermentation conditions of *Sinopodophyllum hexandrum* endophytic fungus on production of podophyllotoxin. Food and Fermentation Industries, 33:28-32.
- Chandra S. (2012). Endophytic fungi: novel sources of anticancer lead molecules. Applied Microbiology and Biotechnology, 95:47–59.
- Channabasava, Govindappa M. (2014). First report of anticancer agent, lapachol producing endophyte, *Aspergillus niger* of *Tabebuia argentea* and its *in vitro* cytotoxicity assays. Bangladesh Journal of Pharmacology, 2014; 9: 129-139.
- Chen J., Qiu X., Wang R., Duan L., Chen S., Luo J., Kong L. (2009). Inhibition of human gastric carcinoma cell growth *in vitro* and *in vivo* by cladosporol isolated from the paclitaxel-producing strain *Alternaria alternata* var. *monosporus*. Biological and Pharmaceutical Bulletin, 32:2072–2074.
- Chowdhary K, Kaushik N. (2015). Fungal endophyte diversity and bioactivity in the Indian medicinal plant *Ocimum sanctum* Linn. PLoS ONE, 10(11): e0141444.
- Cosoveanu A., Hernander M., Iacomi-Vasilescu B., zhang X., Shu S., Wang M., Cabrera R. (2016). Fungi as endophytes in Chinese Artemissia sp. juxtaposed elements of phylogeny, diversity and bioactivity. Mycosphere, 7(2): 102-117.
- Cos P., Maes L., Vanden B.D., Hermans N., Pieters L., Vlietinck A. (2004). Plant substances as anti-HIV agents selected according to their putative mechanism of action. Journal of Natural Products, 67: 284-293.
- Cota B.B., Rosa L.H., Caligiorne R.B., Rabello A.L.T., Alves T.M.A., Rosa C.A., Zani C.L. (2008). Altenusin, a biphenyl isolated from the endophytic fungus *Alternaria* sp., inhibits trypanothione reductase from *Trypanosoma cruzi*. FEMS Microbiology Letters, 285:177–182.
- Cragg G.M., Newman D.J. (2005). Plants as a source of anticancer agents. Journal of Ethnopharmacology, 100:72-79.
- De Clercq E. (2000). Current lead natural products for the chemotherapy of human immunodeficiency virus (HIV) infection. Medicinal Research Reviews, 20: 323-349.
- de Hoog G.S., Horré R. (2002). Molecular taxonomy of the *Alternaria* and *Ulocladium* species from humans and their identification in the routine laboratory. Mycoses, 45: 259–276.
- Ding G., Song Y.C., Chen J.R., Xu C., Ge H.M., Wang X.T., Tan R.X., Chaetoglobosin U. (2006). A cytochalasan alkaloid from endophytic *Chaetomium globosum* IFB-E019. Journal of Natural Products, 69: 302–304.
- Dingle J., McGee P.A. (2003). Some endophytic fungi reduce the density of pustules of *Puccinia recondita* f. sp. *tritici* in wheat. Mycological Research, 107:310–316.
- Duan L. (2009). Isolation and identification of producing endophytic fungi of berberine from the plant *Phellodendrona murense*. Journal of Anhui Agriculture Science, 22: 007.
- Dziezak J.D. (1986). Antioxidants. Food Technology Journal, 40:94.
- Elliott J.A. (1917). Taxonomic characters of the genera *Alternaria* and *Macrosporium*. American Journal of Botany, 4: 439–476.
- Fang Z.F., Yu S.S., Zhou W.Q., Chen X.G., Ma S.G., Li Y., Qu J. (2012). A new isocoumarin from metabolites of the endophytic fungus *Alternaria tenuissima* (Nee & T. Nee: Fr) Wiltshire. Chinese Chemical Letteers, 23:317–320.
- Feng C., Ma Y. (2010). Isolation and anti-phytopathogenic activity of secondary metabolites from *Alternaria* sp. FL25, an endophytic fungus in *Ficus carica*. Chinese Journal of Applied Environmental Biology, 16:76–78.
- Fernandes M.R.V., de Silva T.A.C., Pfenning L.H., da Costa-Neto C.M., Heinrich T.A., de Alencar S.M., de lima M.A., Ikegaki M. (2009). Biological activities of the fermentation extract of the endophytic fungus *Alternaria alternata* isolated from *Coffea arabica* L. Brazillian Journal of Pharmaceutical Sciences, 45(4): 677-685.
- Fries E.M. (1832). Systema mycologicum. vol. 3. E. Moritz, Greifswald, Germany.

- Fritts M., Crawford C.C., Quibell D., Gupta A., Jonas W.B. (2008). <u>Traditional Indian medicine and</u> homeopathy for HIV/AIDS: a review of the literature. AIDS Reseasch Therapy, 5: 25.
- Gangwar M., Verma V.C., Gautam M.K., Nath G. (2015). Isolation and evaluation of antimicrobial activities of endophytic fungal extract from *Mallotus philippinensis* Muell. Applied Microbiology Open Access, 1:103.
- Gao S.S., Li X.M., Wang B.G. (2009). Perylene derivatives produced by *Alternaria alternata*, an endophytic fungus isolated from *Laurencia* species. Natural Product Communication, 4:1477–1480.
- Gong B., Yao X.H., Zhang Y.Q., Fang H.Y., Pang T.C., Dong Q.L. (2015). A cultured endophytic community is associated with plant *Clerodendrum inerme* and antifungal activity. Genetic and Molecular Research, 14(2): 6084-6093.
- Govindappa M., Channabasava R., Sowmya D.V., Meenakshi J., Shreevidya M.R., Lavanya A., Sadananda T.S. (2011). Phytochemical screening, antimicrobial and *in vitro* anti-inflammatory activity of endophytic extracts from *Loranthus* sp. Pharmacognosy Journal, 3(25): 82-90.
- Govindappa M., Sadananda T.S., Channabasava, Ramachandra Y.L., Chandrappa C.P., Padmalatha R.S., Prasad S.K. (2015). *In vitro* and *in vivo* antidiabetic activity of lectin (N-acetyl-galactosamine, 64 kDa) isolated from endophytic fungi, *Alternaria* species from *Viscum album* on alloxan induced diabetic rats. Integrative Obesity Diabetes, 1(1): 11-19.
- Gunasekharan S., Sathiavelu M., Arunachalam S. (2017). *In vitro* antioxidant and antibacterial activity of endophytic fungi isolated from *Mussaenda luteola*. Journal of Appled Pharmaceutical Sciences, 7(8): 234-238.
- Guo B.H., Wang Y.C., Zhou X.W., Hu K., Tan F., Miao Z.Q. (2009). Anendophytictaxol-producing fungus BT2 isolated from *Taxus chinensis* var. *mairei*. African Journal of Biotechnology, 5: 875–877.
- Guo B., Kunming L.H. (1998). A middle vinblastine fungi isolated. Journal of Yunnan University, 20: 214–215.
- Halliwell B. (1994). Antioxidants sense or speculation. Nutrition Today, 29:15–19.
- Harnett S.M., Oosthuizen V., va de Venter M. (2005). Anti-HIV activities of organic and aqueous of *Sutherlandia frutescens* and *Lobostemon trignus*. Journal of Ethnopharmacology, 96: 113-119.
- Hellwig V., Grothe T., Mayer-Bartschmid A., Endermann R., Geschke F.U., Henkel T., Stadler M. (2002). Altersin, a new antibiotic from cultures of endophytic *Alternaria* spp. taxonomy, fermentation, isolation, structure elucidation and biological activities. Journal of Antibiotics, 55: 881-892.
- Hong S.G., Cramer R.A., Lawrence C.B., Pryor B.M. (2005). Alt a 1 allergen homologs from *Alternaria* and related taxa: analysis of phylogenetic content and secondary structure. Fungal Genetics and Biology, 42: 119–129.
- Huang C.H., Pan J.H., Chen B., Yu M., Huang H.B., Zhu X., Lu Y.J., She Z.G., Lin Y.C. (2011). Three bianthraquinone derivatives from the mangrove endophytic fungus *Alternaria* species ZJ9-6B from the South China Sea. Marine Drugs, 9: 832-843.
- Hudson J.B., Imperial V., Haugland R.P., Diwu Z. (1997). Antiviral activities of photoactive perylenequinones. Photochem Photobiol, 65:352–354
- Inderbitzin P., Shoemaker R.A., O'Neill N.R., Turgeon B.G., Berbee M.L. (2006). Systematics and mating systems of two fungal pathogens of opium poppy: the heterothallic *Crivellia papaveracea* with a *Brachycladium penicillatum* asexual state and a homothallic species with a *Brachycladium papaveris* asexual state. Canadian Journal of Botany, 84: 1304–1326.
- Ismaiel A.A., Ahmed A.S., Hassan I.A., El-Sayed R., Al-Zahraa A., El-Din K. (2017). Production of paclitaxel with anticancer activity by two local fungal endophytes, *Aspergillus fumigatus* and *Alternaria tenuissima*. Applied Microbiology and Biotechnology, 101(14): 5831-5846.
- Javanmardi J., Stushnoff C., Locke E., Vivaco J.M. (2003). Antioxidant activity and total phenolic content of Iranian *Ocimum* accessions. Food Chemistry, 83:547-550.
- Jeon Y.T., Ryu K.H., Kang M.K., Park S.H., Yun H., Q T.P., Kim S.U. (2010). Alternariol monomethyl ether and α,β-dehydrocurvularin from endophytic fungi *Alternaria* spp. Inhibit aspersorium formation of *Magnaporthe grisea*. Journal of Korean Society Applied Biological Chemistry, 53: 39–42.

Joly P. (1964). Le genre Alternaria. Encyclopédie mycologique XXXIII, P. Lechevalier, Paris, France.

- Joseph B., Jini D. (2011). Insight into the hypoglycaemic effect of traditional Indian herbs used in the treatment of diabetes. Research Journal of Medicinal Plants, 5: 352-376.
- Kalyanasundaram K., Nagamuthu J., Muthukumaraswamy S. (2015). Antimicrobial activity of endophytic fungi isolated and identified from salt marsh plant in Vellar Estuary. Journal of Microbiology and Antimicrobials, 7(2): 13-20.
- Kasabana S., Hemini S. (1998). Medicinal herb index in Indonesia, Bogor, Indonesia. P.T. Eisai Indonesia. pp. 1-2.
- Kaul S., Wani M., Dhar K.L., Dhar M.K. (2008). Production and GC-MS trace analysis of methyl eugenol form endophytic isolate of *Alternaria* from rose. Annals of Microbiology, 58: 443–445.
- Kjer J., Wray V., Edrada-Ebel R., Ebel R., Pretsch A., Lin W.H., Proksch P. (2009). Xanalteric acids I and II and related phenolic compounds from an endophytic *Alternaria* sp. isolated from the mangrove plant *Sonneratia alba*. Journal of Natural Products, (72):2053–2057.
- Kim S.U., Strobel G.A., Ford E. (1999). Screening of taxol-producing endophytic fungi from *Ginkgo biloba* and *Taxus cuspidata* in Korea. Agricultural Chemistry and Biotechnology, 42: 97-99.
- Kumar S., Kaushik N., Edrada-Ebel R.A., Ebel R., Proksch P. (2011). Isolation, characterization and bioactivity of endophytic fungi of *Tylophora indica*. World Journal of Microbiology Biotechnology, 27(3): 571-577.
- Lawrence D.P., Gannibal P.B., Peever T.L., Pryor B.M. (2012). The sections of *Alternaria*: Formalizing species-groups concepts. Mycologia, 105: 530–546.
- Li W.K., Zhou J.Y., Lin Z.W., Hu Z. (2007). Study on fermentation condition for production of huperzine A from endophytic fungus 2F09P03B of *Huperziaserrata*. Chinese Journal of Medicinal Chemistry, 2: 254–259.
- Li P., Wu Z., Liu T., Wang Y. (2016a). Biodiversity, phylogeny, and antifungal functions of endophytic fungi associated with *Zanthoxylum bungeanum* academic editors: Marcello Iriti and Jianhua Zhu, International Journal of Molecular Science, 17:1541 (8-9).
- Li W., Xu J., Li F., Li C. (2016b). A new antifungal isocoumarin from the endophytic fungus *Trichoderma* sp 09 of *Myoporum bontiodes* A Gray. Pharmacognosy Magazine, 12(48): 259-261.
- Liu Y., Wang M.W. (2008). Botanical Drugs: Challenges and Opportunities: Contribution to Linnaeus Memorial Symposium. Life Science, 82: 445-449
- Lou J., Yu R., Wang X., Mao Z., Fu L., Liu Y., Zhou L. (2016). Alternariol 9-methyl ether from the endophytic fungus *Alternaria* sp. Samif01 and its bioactivities, Brazilian Journal of Microbiology, 47:96–101.
- Lu L., He J., Yu X., Li G., Zhang X. (2006). Studies on isolation and identification of endophytic fungi strain SC13 from pharmaceutical plant *Sabina vulgaris* Ant. and metabolites. Acta Agriculturae Boreali-occidentalis Sinica, 15: 85-89.
- Luo Z.P., Lin H.Y., Ding W.B., He H.L., You-Zhi L. (2015). Phylogenetic diversity and antifungal activity of endophytic fungi associated with *Tephrosia purpurea*. Mycobiology, 43(4):435-443.
- Mani V.M., Soundari A.P.G., Karthiyaini D., Preeth K. (2015). Bioprospecting endophytic fungi and their metabolites from medicinal tree *Aegle marmelos* in Western Ghats, India. Microbiology, 43(3): 303-310.
- Matook S.M. (2005). Antioxidant activities of water-soluble polysaccharides from Buncan (citrus grandis osbeck) fruit flavedo tissues. Pakistan Journal of Biological Science, 8:1472–1477.
- Mesnah L., Viotte G., Sumereau E., Morin J.P., Fillastre J.P. 1987. Haematotoxicity of doxorubicin and 1-(2-chloroethtl)-3cyclohexyl-1-nitrosurea (CCNU) and of their association in rats. Drugs under Experimental and Clinical Research, 13(10): 593-599.
- Michalczyk A., Cieniecka-Roslonkiewicz A., Cholewinska M. (2014). Plant endophytic fungi as a source of paclitaxel. Keria Polonica, 60(4):22-33.
- Monali G., Desale, Bodhankar M.G. (2013). Antimicrobial activity of endophytic fungi isolated from *Vitex negundo* Linn. International Journal of Current Microbiology and Applied Science, 2(12):389-395.

- Musetti R., Polizzotto R., Vecchione A., Borselli S., Zulini L., D'Ambrosio M., Di Toppi L.S., Pertot I. (2007). Antifungal activity of diketopiperazines extracted from *Alternaria alternata* against *Plasmopara viticola*: an ultrastructural study. Micron, 38:643–650.
- Ng T.B., Huang B., Fong W.P., Yeung H.W. (1997). Anti-human immunodeficiency (anti-HIV) natural products with special emphasis on HIV reverse transcriptase inhibitors. Life Sciences, 61: 933-949.
- Ogbonnia S.O., Odimegu J.I., Enwuru V.N. (2008). Evaluation of hypoglycemic and hypolipidemic effects of ethanolic extracts of *Treculia africana* Decne and *Bryopyllum pinnatum* Lam. and their mixture on streptozotocin (STZ)- induced diabetic rats. African Journal of Biotechnology, 7(15):2535–2539.
- Owen N.L., Hundley N. (2004). Endophytes-the chemical synthesizers inside plants. Science Programme, 87:79–99.
- Park J.H., Choi G.J., Lee H.B., Kim K.M., Jung H.S., Lee S.W., Jang K.S., Cho K.Y., Kim J.C. (2005). Griseofulvin from *Xylaria* sp. strain F0010, and endophytic fungus of *Abies holophylla* and its antifungal activity against plant pathogenic fungi. Journal of Microbiology and Biotechnology, 15:112–117.
- Patel D.K., Prasad S.K., Kumar R., Hemelatha S. (2012). An overview on antidiabetic medicinal plants having insulin mimetic property. Asian Pacific Journal of Tropical Biomedicine, 2:320–330.
- Phongpaichit S., Nikom J., Rungjindamai N., Sakayaroj J., Hutadilok-Towatana N., Rukachaisirikul V., Kirtikara K. (2007). Biological activities of extracts from endophytic fungi isolated from *Garcinia* plants. FEMS Immunology, Medicinal Microbiology, 51: 517–525.
- Pietta P., Simonetti P., Mauri P. (1998). Antioxidant activity of selected medicinal plants. Journal of Agriculture Food Chemistry, 46: 4487-4490.
- Porras-Alfaro A., Bayman P. (2011). Hidden fungi, emergent properties: endophytes and microbiomes. Annual Review of Phytopathology. 49: 291-315.
- Preuss C.G.T. (1851). Übersicht untersuchter pilze, besonders aus der Umgegend von Hoyerswerda. Linnaea, 24: 99–153.
- Pryor B.M., Bigelow D.M. (2003). Molecular characterization of *Embellisia* and *Nimbya* species and their relationship to *Alternaria*, *Ulocladium* and *Stemphylium*. Mycologia, 95: 1141–1154.
- Pryor B.M., Creamer R., Shoemaker R.A., McLain-Romero J., Hambleton S. (2009). *Undifilum*, a new genus for endophytic *Embellisia oxytropis* and parasitic *Helminthosporium bornmuelleri* on legumes. Botany, 87: 178–194.
- Qin J.C., Zhang Y.M., Hu L., Ma Y.T., Gao J.M. (2009). Cytotoxic metabolites produced by *Alternaria* no.28, an endophytic fungus isolated from *Ginkgo biloba*. Natural Products Communication, 4: 1473–1476.
- Raman B.V., Krishna N.V., Rao N.B., Saradhi P.M., Rao B.M.V. (2012). Plants with antidiabetic activities and their medicinal values. International Research Journal of Pharmacology, 3(3):11–15.
- Roopa G., Madhusudhan M.C., Sunil K.C.R., Lisa N., Calvin R., Poornima R., Zeinab N., Kini K.R., Prakash H.S., Geetha N. (2015). Identification of taxol producing endophytic fungi isolated from *Salacia oblonga* through genomic mining approach. Journal of Genetic Engineering and Biotechnology, 13(2): 119-127.
- Runa F., Park M., Pryor B. (2009). *Ulocladium* systematics revisited: phylogeny and taxonomic status. Mycological Progress, 8: 35–47.
- Saccardo P.A. (1886). Sylloge Fungorum Omnium hucusque cognitorum, Volume 4. Padua, Italy.
- Sadananda T.S., Nirupama R., Chaithra K., Govindappa M., Chandrappa C.P., Vinay B.R. (2011). Antimicrobial and antioxidant activities of endophytes from *Tabebuia argentea* and identification of anticancer agent, Lapachol. Journal of Medicinal Plants Research, 5(16):3643-3652.
- Sandhu S.S., Kumar S., Aharwa R.P. (2014a). Isolation and identification of endophytic fungi from *Ricinus communis* Linn. and their antibacterial activity. International Journal of Research Ppharmaceutical Chemistry, 4(3), 611-618.
- Sandhu S.S., Rajak R.C., Shukla H., Aharwal P.R., Kumar S. (2014b). Endophytic fungi: as a source of antimicrobials bioactive compounds. Journal of Pharm Science, 3: 1179-1197.

- Schulz B., Rommert A.K., Dammann U., Aust H.J., Strack D. (1999). The endophyte-host interaction: a balanced antagonism? Mycological Research, 103:1275–1283.
- Selvi B.K., Balagengatharathilagam P. (2014). Isolation and screening of endophytic fungi from medicinal plants of Virudhunagar district for antimicrobial activity. International Journal of Science and Nature, 5(1): 147-155.
- Shaaban M., Shaaban K.A., Abdel-Aziz M.S. (2012). Seven naptho-γ-pyrones from themarine derived fungus *Alternaria alternata*: structure elucidation and biological properties. Organic Medicinal Chemical Letter, 2:6.
- Shahidi F., Wanasundara P.K.J.P.D. (1992). Phenolic antioxidants. Critical Review on Food Science Nutrition, 32: 67-103.
- Sharma A.K., Aggarwal A., Singhal V.K. (2012). Treatment of diabetes mellitus with Indian herbal Drugs. International Journal of Advanced Research in Pharmaceutical and Bio Sciences, 1(2):145–153.
- Shaw J.E., Sicree R.A., Zimmet P.Z. (2010). Global estimates of the prevalence of diabetes for 2010 and 2030. Diabetes Research Clinical Practice, 87:4–14.
- Shi J., Zeng Q., Liu Y., Pan Z. (2012). *Alternaria* sp. MG1, a resveratrol-producing fungus: isolation, identification, and optimal cultivation conditions for resveratrol production. Applied Microbiology Biotechnology, 95: 369–379.
- Shweta S., Gurumurthy B.R., Ravikanth G., Ramanan U.S., Shivanna M.B. (2013). Endophytic fungi from *Miquelia dentata* Bedd., produce the anti-cancer alkaloid, camptothecine. Phytomedicine, 20:337–342.
- Siegel M.R., Latch G.C.M., Bush L.P., Fannin N.F., Rowan D.D., Tapper B.A., Bacon C.W., Johnson M.C. (1990). Fungal endophyte-infected grasses: alkaloid accumulation and aphid response. Journal of Chemical Ecology, 16:3301–3315.
- Silva-Hughes A.F., Wedge D.E., Cantrell C.L., Carvalho C.R., Pan Z., Moraes R.M., Madoxx V.L., Rosa L.H. (2015). Diversity and antifungal activity of the endophytic fungi associated with the native medicinal cactus *Opuntia humifusa* (*Cactaceae*) from the United States. Microbiological Research, 175:67-77.
- Simmons E.G. (2007). *Alternaria*. An identification manual. CBS Biodiversity Series 6. CBS Fungal Biodiversity Centre, Utrecht, The Netherlands.
- Singh K., Singh K., Dangi C.B.S. (2017). Endophytic fungi with anti-diabetic activities isolated from amla fruits. International Journal of Research in Applied, Natural and Social Sciences, 5(1): 121-128.
- Singh I.P., Bharate S.B., Bhutani K.K. (2005). <u>Anti-HIV natural products. Current Science</u>, 89: 269-290.
- Soliman S.S.M., Raizada M.N. (2013). Interactions between cohabitating fungi elicit synthesis of taxol from an endophytic fungus in hostTaxus plants. Fronteniers in Microbiology, 4:1-14.
- Srinivasan R.P., Nigam A., Aruna J., Alam M.A., Ishara L., Chamith Y.H., Chikkaswamy B.K. (2015). Antimicrobial activity in cultures of endophytic fungi isolated in some medicinal plant species. International Journal of Advanced Research in IT and Engineering, 4(1): 1-24.
- Staniek A., Woerdenbag H.J., Kayser O. (2010). Screening the endophytic flora of *Wollemina nobilis* for alternative paclitaxel sources. Journal of Plant Interactions, 5(3): 189-195.
- Strobel G.A., Hess W.M., Ford E., Sidhu R.S., Yang X. (1996). Taxol from fungal endophytes and issue of biodiversity. Journal of Industrial Microbiology, 17: 417-423.
- Strobel GA, Dirkse E, Sears J, Markworth C. 2001. Volatile antimicrobials from *Muscador albus*, a novel endophytic fungus. Microbiology, 147:2943–2950.
- Strobel G.A. (2003). Endophytes as sources of bioactive products. Microbial Infection, 5: 535-544.
- Strobel G. (2006). *Muscodor albus* and its biological promise. Journal of Indian Microbiology and Biotechnology, 33:514–522.
- Taware A.S., Rajurkar S.K. (2015). Diversity assessment of endophytic fungi from *Azadirachta indica* A. Juss from various regions of Aurangabad, Maharastra (India). International Journal of Innovative Science, Engineering & Technology, 2(7): 96-107.

- Tenguria R.K., Firodiya A.D. (2016). Cytotoxic activities of endophytic fungi isolated from central region of Madhya Pradesh. Journal of Innovations in Pharmaceuticals and Biological Sciences, 3(3): 29-38.
- Tian R., Yang Q., Zhou G., Tan J., Zhang L., Fang C. (2006). Taxonomic study on a taxol producing fungus isolated from bark of *Taxus chinensis* var. *mairei*. Journal of Wuhan Botanical Research, 24: 541-545.
- Tian J., Fu L., Zhang Z., Dong X., Xu D., Mao Z., Liu Y., Lai D. (2017). Dibenzo-α-pyrones from the endophytic fungus *Alternaria* species Sami01: isolation, structure elucidation, and their antibacterial and antioxidant activities. Natural Product Research, 31(4): 387-396.
- Umashankar T., Govindappa M., Ramachandra Y.L., Chandrappa C.P., Padmalatha Rai S., Channabasava R. (2015). Isolation, purification and *in vitro* cytotoxicity activities of coumarin isolated from endophytic fungi, *Alternaria* species of *Crotalaria pallida*. Indo American Journal of Phamaceutical Research, 5(2): 926-936.
- Urquiaga I., Leighton F. (2000). Plant polyphenol antioxidants and oxidative stress. Biology Research, 33:9716–9760.
- Vasundhara M., Kumar A., Reddy M.S. (2016). Molecular approaches to screen bioactive compounds from endophytic fungi. <u>Frontiers in Microbiology</u>, 7: 1774.
- Velioglu Y.S., Mazza G., Gao L., Oomah B.D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables and grain products. Journal of Agriculture Food Chemistry, 46: 4113-4117.
- Vlietinck A.J., De Bruyne T., Apers S., Pieters L.A. (1998). Plant derived leading compounds for chemotherapy of human immunodeficiency virus (HIV) infection. Planta Medica, 64: 97-109.
- von Esenbeck C.G.N. (1816). Das system der pilze und schwämme. Wurzburg, Germany.
- von Keissler K. (1912). Zur kenntnis der pilzflora krains. Beihefte zum Botanischen Zentralblatt 29: 395–440.
- Wallroth C.F.W. (1833). Flora Cryptogamica Germaniae Sectio 2. J.L. Schrag, Nürnberg, Germany.
- Wang J.H.W., Nie H.L., Tam S.C., Huang H., Zheng Y.T. (2004). Anti-HIV-1 property of trichosanthin correlates with its ribosome inactivating activity. FEBS Letters, 531(2): 295-298.
- Wang J., Cox D.G., Ding W., Huang G., Lin Y., Li C. (2014). Three new resveratrol derivatives from the Mangrove endophytic fungus *Alternaria* sp. Marine Drugs, 12:2840-2850.
- Wang X.Z., Luo X.H., Xiao J., Zhai M.M., Yuan Y., Zhu Y., Crews P., Yuan C.S., Wu Q.X. (2014). Pyrone derivatives from the endophytic fungus *Alternaria tenuissima* SP-07 of Chinese herbal medicine *Salvia przewalskii*. Fitoterapia, 99:184–190.
- Wang M., Sun Z.H., Chen Y.C., Liu H.X., Li H.H., Tan G.H., Li S.N., Guo X.L., Zhang W.M. (2016). Cytotoxic cochlioquinone derivatives from the endophytic fungus *Bipolaris sorokiniana* derived from *Pogostemon cablin*. Fitoterapia, 110:77-82.
- Wellensiek B.P., Ramakrishnan R., Bashyal B.P., Eason Y., Gunatilaka A.A.L., Ahmad N. (2013). Inhibition of HIV-1 replication by secondary metabolites from endophytic fungi of desert plants. Open Virol Journal, 7: 72-80.
- West C.P. (1994). Physiology and drought tolerance of endophyte— infected grasses. In: Bacon CW, White JF (eds) Biotechnology of endophytic fungi of grasses. CRC Press, Boca Raton, pp 87–99.
- Wiltshire S.P. (1933). The foundation species of *Alternaria* and *Macrosporium*. Transactions of the British Mycological Society, 18: 135–160.
- Wiltshire S.P. (1938). The original and modern conceptions of *Stemphylium*. Transactions of the British Mycological Society, 21: 211–239.
- Wiyakrutta S., Sriubolmas N., Panphut W., Thong N., Danwisetkanjana K., Ruangrungsi N., Meevootisom V. (2004). Endophytic fungi with anti-microbial, anti-cancer and anti-malarial activities isolated from Thai medicinal plants. World Journal of Microbiology and Biotechnology, 20(3):265-272.
- Wu S.H., Chen Y., Li Z., Yang L., Li S. (2011). Metabolites of the endophytic fungus *Alternaria* sp PR14 of *Paeonia delayayi*. Natural Resesearch Development, 23: 850-852.

- Xiao Y., Li H.X., Li C., Wang J.X., Li J., Wang M.H., Ye Y.H. (2013). Antifungal screening of endophytic fungi from *Ginkgo biloba* for discovery of potent anti-phytopathogenic fungicides. FEMS Microbiology Letters, 339(2): 130-135.
- Xia G., Li J., Li H., Long Y., Lin S., Lu Y., He L., Lin Y., Liu L., She Z. (2014). Alterporriol-type dimers from the mangrove endophytic fungus, *Alternaria* sp. (SK11), and their MptpB inhibitions. Marine Drugs, 12:2953-2969.
- Yadav M., Yadav A., Kumar S., Yadav J.P. (2016). Spatial and seasonal influences onculturable endophytic mycobiota associated with different tissues of *Eugenia jambolana* Lam and their antibacterial activity against MDR strains. BMC Microbiology, 16:44.
- Yang X., Guo S., Zhang L., Shao H. (2003). Selection of producing podophyllotoxin endophytic fungi from podophyllin plant. Natural Product Research and Development, 15: 419-422.
- Yang S.S., Gragg G.M., Newman D.J, Bader J.P. (2001). Natural product based anti-HIV drug discovery and development facilitated by the NCI developmental therapeutics program. Journal of Natural Products, 64: 265-277.
- Yuan L., Zhao P.J.M., Ma J., Li G.H., Shen Y.M. (2008). Tricycloalternarenes A-E: five new mixed terpenoids from the endophytic fungal strain *Alternaria alternata* Ly83. Helv Chim Acta, 91:1588-1594.
- Zhang L.Q., Guo B., Li H.Y. (1998). Isolation of an fungus producing vinbrastine. Journal of Yunnan University (Natural Science), 20: 214–215.
- Zhang N., Zhang C., Xiao X., Zhang Q., Huang B. (2016). New cytotoxic compounds of endophytic fungus *Alternaria* species isolated from *Broussoneta papyrifera* (L.) Vent. Fitoterapia, 110: 173-180.
- Zhao J., Zhou L., Wang J., Shan T., Zhong L., Liu X., Gao X. (2012). Endophytic fungi for producing bioactive compounds originally from their host plants. In: Mendez-Vilas A, editor. Current Research, Technology and Education Topics in Applied Microbiology and Microbial Biotechnology. Vol. 1. Formatex Research Center; Badajoz, Spain: 2012. pp. 567–576.
- Zheng L., Ma Y., Zhang Y. (2014). *In vitro* α-glucosidase inhibitory activity of endophytic *Alternaria* sp. S8 isolated from *Morus alba* L. Planta Medica, 80: PIN17.
- Zhou X., Wang Z., Jiang K., Wei Y., Lin J., Sun X. (2007). Screening of taxol-producing endophytic fungi from *Taxus chinensis* var. *mairei*. Prikl. Biokhim.Mikrobiology, 43:490–494.
- Zou W.X., Meng J.C., Lu H., Chen G.X., Shi G.X., Zhang T.Y., Tan R.X. (2000). Metabolites of *Colletotrichum gloeosporioides*, an endophytic fungus in *Artemisia mongolica*. Journal of Natural Products, 63:1529–1530.

Table 1. in supplementary file.